

Anatoxin-a, Cylindrospermopsin, Microcystins/Nodularins, & Saxitoxins Report

Project: Washoe County Community Services Department

Submitted to: John Hulett

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Sample Receipt Date: 21 September 18 Sample Condition: 7.6 °C upon arrival

Report# 180920_Washoe County Community Services Department

Date Prepared: 24 September 18 Prepared by: Kamil Cieslik

Sample ID	Site/Description	Collection Date	Collection Time
1	Lemmon Lake Inlet	20 September 18	1200

Analytes: Anatoxin-a (ANTX-A), Cylindrospermopsin (CYN), Microcystins/Nodularins (MCs/NODs), Saxitoxin (STX/PSTs)

Sample Preparation

Water Sample Ultrasonication

The sample was received and inverted for 60 seconds to mix. A subset of the sample was sonicated to release toxins and prepared for analyses.





Analytical Techniques

Enzyme-Linked Immunosorbent Assay (ELISA)

MCs/NODs

A microcystins/nodularins Adda ELISA (Abraxis) was utilized for the quantitative and sensitive congener-independent detection of MCs/NODs (US EPA Method 546 & Ohio EPA DES 701.0). The current method reporting limit is 0.15 ng/mL (ppb) based on kit sensitivity, dilution factors and initial demonstration of capability.

STX

A saxitoxin specific ELISA (Abraxis PN 52255B) was utilized for the detection and quantification of saxitoxin and related analogs (paralytic shellfish toxins – PSTs). The current method reporting limit is 0.05 ng/mL (ppb) based on kit sensitivity and dilution factors. Based on manufacture instructions, the STX ELISA is less cross-reactive to other PSTs and will likely underestimate total PSTs/Saxitoxins. Reported cross-reactivities are as follows: NEO (1.3%), dcSTX (29%), GTX2/3 (23%), GTX5 (23%), dcGTX2/3 (1.4%), dcNEO (0.6%) & GTX1/4 (<0.2%).

Liquid chromatography mass spectrometry/mass spectrometry (LC-MS/MS)

ANTX-A

The $[M+H]^+$ ion for ANTX-A (m/z 166) was fragmented and the product ions (m/z 91, 131 & 149) were monitored. The sample response was compared to the LFSM response utilizing the quantification ion, m/z 91. An external standard curve was used to determine LFSM returns.

CYN

The $[M+H]^+$ ion for CYN (m/z 416) was fragmented and the product ions (m/z 194, 274, 336) were monitored. The sample response was compared to the LFSM response utilizing the quantification ion, m/z 336. An external standard curve was used to determine LFSM returns.





Quality Control

Table 1: LFSM QC samples prepared for analyses.

Analyte	Concentration (ng/mL)	Sample ID	QC Type	Return
MC-LR	1.0	1 (Lemmon Lake Inlet)	LFSM	84%
CYN	0.1	1 (Lemmon Lake Inlet)	LFSM	82%
ANTX-A	0.1	1 (Lemmon Lake Inlet)	LFSM	107%
STX	0.2	1 (Lemmon Lake Inlet)	LFSM	80%

Additional Quality Control/Quality Assurance checks included method blanks, LFBs, and standard curves.

Flag
Analytical result is estimated due to ineffective quenching.
Analyte was positively identified; the associated numerical value is estimated.
The reported result is estimated because the sample was not analyzed within required holding time.
Analytical result is estimated. Analyte was detected in associated reagent blank as well as the samples.
Analytical result is estimated. Values achieved were outside calibration range.
Spiked sample control was outside limits
The reported result is estimated because the sample exceeded temperature threshold when received

Abbreviations					
NA	Not Applicable	LFSM	Lab Fortified Sample Matrix		
MDL	Method Detection Limit	LFSMD	Lab Fortified Sample Matrix Duplicate		
MQL	Method Quantification Limit	LD	Lab Duplicate		
ND	Not Detected above the MDL	IS	Internal Standard		
Blank	Regent Water free from interferences	_	Not Analyzed		
LFB	Lab Fortified Blank	MRL	Method Reporting Limit		



Summary of Results

Sample ID	MCs/NODs (ng/mL)	CYN (ng/mL)	ANTX-A (ng/mL)	STX (ng/mL)
1 (Lemmon Lake Inlet)	ND	ND	ND	ND
MRL (ng/mL)	0.15	0.05	0.05	0.05
Analyst Initials	KC	MA	MA	KC
Date Analyzed	9/24/18	9/21/18	9/21/18	9/21/18

Submitted by:

Mark T. Aubel, Ph.D.

Date:

September 24, 2018

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